# Molecular characterisation of biogenic secondary organic aerosols using ion mobility spectrometry - quadrupole time-of-flight mass spectrometry

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# Introduction

Molecular characterisation of unknown organic compounds is challenging, in particular the molecular identification of secondary organic aerosol constituents is extremely demanding due to their low quantities and high complexity of the sample matrix. The presence of multiple structural isomers and their complex structures often make the synthesis of reference compounds very difficult, hindering a positive identification of these compounds.

Traditionally, chromatographic separation is coupled to mass spectrometry for the structural elucidation of unknown compounds in atmospheric aerosol samples. Chromatographic separation provides information on the volatility or polarity of the unknowns and mass spectrometry provides their structural and/or molecular mass information. In particular, the applications of derivatisation GC/MS and HPLC/ESI-MS techniques in recent years brought significant advances in our understanding of SOA constituent structures, leading to the identification of methyltetrols from isoprene oxidation (Claeys et al., 2004), tricarboxylic acid from apinene oxidation (Szmigielski et al., 2007), and structural proposals for various organosul-Iphates (Gómez-González et al., 2008; linuma et al., 2007ab, 2009; Surratt et al., 2007, 2008) Ion mobility spectrometry coupled to mass spectrometry (IMS-MS) is an analytical tool which offers a rapid separation of molecules by their mobilities in the gas phase, hence their collision cross-sections (CCS). The CCSs of the ions can be calculated through the measurement of ion mobilities, and structural information of the ions can be obtained by comparison with computer model estimated CCSs. Here we provide the results from the first application of IMS-TOFMS on molecular characterisation of monoterpene originating organosulfates and nitrooxy-organosulfates.

# Results and discussion

## a-Pinene originating organosulfates







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# Methods

### Instrumentation

Samples were analysed using an ultra performance liquid chromatography coupled to electrospray ionisation ion mobility spectrometry and quadrupole time-of-flight mass spectrometry (UPLC/(-)ESI-IMS-QTOFMS, Waters Synapt HDMS). Figures 1 and 2 show schematic diagrams of the IMS-QTOFMS used in this study.

The drift times of ions in the DETECTOR TRIWAVE DRE LENS IMS are linearly related to their ION GUIDE QUADRUPOLE mobilities. The mobility (K) of a ANALYTE SPRAY

b-Pinene originating organosulfates



Figure 5 shows an EIC, drift times and fragmentation patterns for b-pinene originating organosulfates ( $C_{10}H_{17}O_5S$ , m/z 249.0802) reported in Surratt et al. (2008) and linuma et al. (2007a, 2009). The peaks 1 is assigned to a primary carbon substituted organosulfate and the peak 2 is assigned to a tertiary carbon substituted isomer based on the fragmentation data. Measured CCSs are somewhat smaller than the estimated CCSs for the peak 1. This likely originates from an H-bond between a sulfate and a hydroxyl group as the inclusion of H-bond in the model structure makes the estimated CCS values closer to the measured CCS.

gas phase ion is defined as K = $v_{\rm D}/E$  where  $v_{\rm D}$  and E are the drift velocity and the electric field, respectively. The mobility is related to CCS as follows:

#### where

16

q: ion charge m: mass of ion, m<sub>B</sub>: mass of buffer gas k: Boltzmann constant From Ion Source T: temperature N: buffer gas number density  $\Omega$ : collision cross section  $K = \frac{\sqrt{18\pi}}{2}$  $m_B \sqrt{kT} N\Omega$ Electric Field Vm

When the accurate T and N cannot be measured, ions of known CCSs can be used to calibrate the drift time measurements (Ruotolo et al., 2008). In the present study, CCS values available for polyamino acids (Wyttenbach et al., 1998; Henderson et al., 1999) are used for calibration of the the instrument. Figure shows





#### Figure 2. Separation of gas phase ions in a drift tube

a calibration curve used in this study. The correlation coefficient,  $R^2 > 0.99$ , is sufficiently higher than the recommended value of 0.98 in Ruotolo et al. (2008).

## Molecular modelling



Figure 5. EIC (top), drift times (middle) and fragmentation data (bottom) for m/z 249.

# Nitrooxy-organosulfates (NOS) originating from monoterpenes



100-

100-

Peak 5

100-

Figure 6 shows an EIC, drift times and fragmentation patterns for NOSs originating from monoterpenes (C<sub>10</sub>H<sub>16</sub>NO<sub>7</sub>S<sup>-</sup>, m/z 294.0653) reported in linuma et al. (2007a) and Surratt et al. (2008). Only a-pinene originating NOSs are compared in this study as no other monoterpenes are reported to produce atmospherically relevant NOSs so far (Surratt et al., 2008). Six peaks are observed in the EIC. Peaks 1-5 produce SO<sub>4</sub> radical, suggesting a sulfate group at a tertiary carbon atom for these peaks (tNOS). Estimated CCSs for tNOS agree well with measured CCSs for the peaks 1-5. The measured CCS for the peak 4 is somewhat smaller than the others, inferring a Z isomer structure for this peak. Measured and estimated CCSs for NOS with a sulfate group at a secondary carbon atom (sNOS) did not agree well (peak 6). Further study is needed to elucidate the structure of this peak.





## 200 DT' Figure 3. W' as a function of DT'. Gly = glycine. Ala = Alanine.

#### Reduced mass $\Omega' = CCS \times \Omega$

DT '= Measureddrift time- travelling time in TOFMS Note that reported CCSs are for protonated forms. These values are corrected for de-protonated forms in the pre-Differences in protonated and deprotonated CCSs are smaller than 1%

Detailed protocol for CCS estimation reported by Ruotolo et al. (2008) is used in the present study. Briefly, CCSs are estimated using open source software MOBCAL (Mesleh et al., 1996). A molecular coordinate file that contains a structure

of target compound is created using Maestro software (Schrödinger, http://www.schrodinger.com/). A trajectory method is used to estimate CCSs.

				0-+
				Peak 6
				100 g 96.9612
	M. Claeys et al. (2004). Science, 303, 1173.	M. F. Mesleh et al. (1996). J. Phys. Chem. 100, 16082		61.9899
References	Y. Gómez-González et al. (2008). J. Mass Spectrom., 43, 371.	J. D. Surratt et al. (2007), Environ. Sci. Technol., 41, 517.	•	≈ 79.9599 231.0665
	S. C. Henderson et al. (1999). J. Phys. Chem. B, 103, 8780.	J. D. Surratt et al. (2008). J. Phys. Chem. A, 112, 8345-8378, 10.1021/jp802310p		
	Y. linuma et al. (2007a). Environ. Sci. Technol., 41, 6678.	B. Ruotolo et al. (2008). Nature. doi:10.1038/nprot.2008.78		• 0 <sup>-11</sup>
	Y. linuma et al. (2007b). Atmos. Environ., 41, 5571.	R. Szmigielski et al. (2007). Geophys. Res. Lett., 34, doi:10.1029/2007GL031338		Figure 6 EIC (top) drift times (middle) and fragm
	Y. Iinuma et al. (2009). Phys. Chem. Chem. Phys.,	T. Wyttenbach et al. (1998). J. Am. Chem. Soc., 120, 5098.	•	Figure 6. EIC (top), drift times (midule) and haging
	doi:10.1039/b904025k	ч ч маата да и шалдаата хил и муала на тручка та жал тал. ч тр		tation data (bottom) for m/z 294.